

Fatty Acid Composition and Nutritional Indices/Ratios of Colostrum and Milk from Murrah and “Murrah × Carabao” Crossbred Buffaloes

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This study aims to determine the fatty acid (FA) composition and compare FA-based nutritional indices/ratios of colostrum and milk obtained on the 30th, 60th, and 90th day of lactation from 27 Murrah and 18 “Murrah × Carabao” crossbred buffaloes. The major saturated FA (SFA) in colostrum and milk is palmitic acid (C16:0), comprising 32–33% by weight of total FAs. Other important SFAs – myristic acid (C14:0), stearic acid (C18:0), and lauric acid (C12:0) – were lower ($P < 0.05$) in colostrum than in milk. Oleic acid (C18:1-n9c), a monounsaturated fatty acid (MUFA), is the second most abundant FA in colostrum (28.5%) and milk (18.0–18.8%). Arachidonic acid (C20:4-n6, AA) is the dominant polyunsaturated fatty acid (PUFA) in colostrum (0.92%) and milk (0.42–0.45%). Conjugated linoleic acid (C18:2-c9t11, CLA) was higher in colostrum (0.64%) than in milk (0.14–0.16%). Colostrum and milk had a very low PUFA/SFA ratio (0.02–0.06: 1). The linoleic acid (C18:2-n6, LA) to α -linolenic acid (C18:3-n3, ALA) ratio was higher in colostrum (3.21: 1) than in milk (0.62–1.55: 1). The omega-6 (LA and AA) to omega-3 [ALA and docosahexaenoic acid (C22:6-n3, DHA)] or n-6/n-3 ratio was more balanced for milk (1.76-2.34: 1) than colostrum (3.37: 1). Colostrum had lower atherogenicity (2.53 vs. 4.50–4.66), lower thrombogenicity (2.68 vs. 4.48–4.59), and higher health-promoting index (0.39 vs. 0.21–0.22) than milk. The hypocholesterolemic/hypercholesterolemic (h/H) ratio was higher for colostrum (0.64: 1) than milk (0.34–0.36: 1). Except for AA, the FA composition of colostrum and milk were not significantly different between Murrah and “Murrah × Carabao” crosses ($P > 0.05$).

Keywords: colostrum, fatty acids, milk, Murrah, “Murrah × Carabao” buffaloes, nutritional indices

INTRODUCTION

Milk FAs are important nutritional components that may have positive effects on human health (Cichosz *et al.* 2020). On the other hand, milk and some dairy products (butter) have also been criticized for the unfavorable FA profile in milk fat (Hanus *et al.* 2018). The FA composition is an important measure of the quality of raw milk or colostrum used in the manufacture of dairy products with improved nutritional quality and health benefits available

to consumers (Timlin *et al.* 2021). Recently, FA in colostrum has attracted interest as a potentially beneficial food ingredient or functional food (Mehra *et al.* 2021; Silva Galdino *et al.* 2021). Colostrum yield, however, is only 0.5% of the cow’s annual milk production. Surplus colostrum was even previously thought of as unmarketable since the focus was more on its role in the development of the calf (O’Callaghan *et al.* 2020).

While studies on FAs in buffalo colostrum are scarce, several studies have reported the milk FA composition of milk from buffaloes belonging to different breeds

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such as the Anatolian water buffalo in Turkey (Cinar *et al.* 2019), Bhadawari buffaloes in India (Kushwaha *et al.* 2018), Bulgarian Murrah (Penchev *et al.* 2016; Ilieva *et al.* 2020), Kundi in Pakistan (Talpur *et al.* 2007), Mediterranean buffaloes in Italy (Varricchio *et al.* 2007; Pegolo *et al.* 2017) and in France (Menard *et al.* 2010) – plus Murrah buffaloes in Brazil (Fernandes *et al.* 2007) and Nili-Ravi in Pakistan (Talpur *et al.* 2007; Qureshi *et al.* 2015). However, the reports listed above vary in the number of individual FAs involving different or no measures of nutritional quality that are indicative of the effect of FAs on human health and disease (Chen and Liu 2020).

In the Philippines, buffalo milk is the second most-produced milk after cow's (bovine) milk, contributing about one-third of the 2019 dairy output of 24.4 million liters (PSA 2020). Unfortunately, very limited work has been carried out to investigate the FA profile of buffalo colostrum and milk. In this regard, the objective of this study was to compare the FA composition and FA-based nutritional indices of colostrum and milk obtained on the 30th, 60th, and 90th day of lactation from Murrah and "Murrah × Carabao" crossbred buffaloes.

MATERIALS AND METHODS

This study was conducted in compliance with the requirements of the Institutional Animal Care and Use Committee of the University of the Philippines Los Baños (UPLB), Laguna, Philippines.

Experimental Animals and Data

Forty-five (45) buffaloes (*i.e.* 27 Murrah and 18 "Murrah × Carabao" crosses) that calved between 04 Apr 2019 and 27 Jul 2020 at the Philippine Carabao Center (PCC) dairy farm in UPLB, Laguna, Philippines were used in the study. The crossbred buffaloes consisted of 3 "50% Murrah-50% Carabao," 3 "75% Murrah-25% Carabao," 9 "87.5% Murrah-12.5% Carabao," and 3 "93.75% Murrah-6.25% Carabao" (preliminary analyses of colostrum and milk showed no significant differences in milk composition among the buffalo crosses; data not shown). The age at calving and parity of the experimental animals was 7.83 ± 3.08 yr and 4.07 ± 2.44 calving, respectively. Buffaloes were equally managed to fulfill all welfare requirements and were kept in individual parturition pens about 2 wk before calving. In addition to forage, the buffaloes were fed with commercial lactating feed concentrates containing 15.52% crude protein, 5.87% crude fat, 11.99% crude fiber, 7.90% moisture, 8.74% ash, 1.02% calcium, and 0.58% phosphorus.

Four milk samples were collected from each buffalo consisting of colostrum taken on the day of calving, and subsequent milk on the 30th, 60th, and 90th day post parturition, placed in plastic bottles, and immediately frozen at -20 °C until further analysis. The number and distribution of buffaloes and colostrum/milk samples collected by breed are shown in Table 1. Other data were also collected including age at calving, parity (number of calving), test-day milk yield, and fat percentage corresponding with each colostrum or milk sample.

A total of 19 medium- to very long-chained FAs were each analyzed as a percentage (g/ 100 g) of total FAs in colostrum and milk samples, *i.e.* eight SFAs [C12:0 (lauric acid), C14:0 (myristic acid), C15:0 (pentadecylic acid), C16:0 (palmitic acid), C17:0 (margaric acid), C18:0 (stearic acid), C20:0 (arachidic acid), and C22:0 (behenic acid)], six MUFAs [C14:1 n-5 (myristoleic acid), C16:1 n-7 (palmitoleic acid), C18:1 n-9 (oleic acid), C18:1 n-7 (trans-vaccenic acid), C20:1 n-11 (eicosenoic acid), and C22:1 n-9 (erucic acid)], and five PUFAs [C18:2 c9tll (CLA), C18:2 n-6 (LA), C18:3 n-3 (ALA), C20:4 n-6 (AA), and C22:6 n-3 (DHA)].

The FA composition of feed concentrates was also analyzed and found to contain 64.32% SFA [C12:0 (37.64%), C14:0 (13.60%), C16:0 (10.02%), C18:0 (2.48%), C20:0 (0.50%), and C22:0 (0.08%)], 14.24% MUFA [C16:1 n-7 (0.07%) and C18:1n9c (14.17%)], and 8.12% PUFA [C18:2 n-6 (8.00%) and C18:3 n-3 (0.12%)].

Following Chen and Liu (2020), six FA groups (*i.e.* SFA, MUFA, PUFA, UFA = MUFA + PUFA, omega-3 FA = C18:3 n-3 and C22:6 n-3, and omega-6 FA = C18:2 n-6 + C20:4 n-6) and seven FA-based nutritional indices/ratios with health implications were also determined. The FA-based nutritional indices/ratios were: 1) PUFA/SFA ratio, 2) n-6/n-3 ratio, 3) LA/ALA ratio, 4) atherogenicity index (IA), 5) thrombogenicity index (IT), 6) health-promoting index (HPI), and 7) h/H ratio.

The IA and IT were calculated using the equations of Ulbricht and Southgate (1991) as follows:

$$IA = [C12:0 + (4 \times C14:0) + C16:0] / \Sigma UFA \quad (1)$$

$$IT = (C14:0 + C16:0 + C18:0) / [(0.5 \times MUFA) + (0.5 \times n - 6 PUFA) + (3 \times n - 3) + (n - 3/n - 6)] \quad (2)$$

The HPI used by Chen *et al.* (2004) was:

$$HPI = UFA / [C12:0 + (4 \times C14:0) + C16:0] \quad (3)$$

The h/H ratio used by Mierlita (2018) was:

Table 1. Number of samples and least squares means of colostrum or test-day milk yield, fat content, and daily fat yield for different breeds and different milk types.

| | Murrah | “Murrah × Carabao” crosses | Total |
|---------------------------|---------------------------|----------------------------|---------------------------|
| No. of buffaloes | 27 | 18 | 45 |
| No. of samples | | | |
| - Colostrum | 27 | 18 | 45 |
| - Milk 30-d | 26 | 17 | 43 |
| - Milk 60-d | 25 | 17 | 42 |
| - Milk 90-d | 22 | 16 | 38 |
| Total no. of samples | 100 | 68 | 168 |
| Yield, kg | | | |
| - Colostrum | 3.97 ± 0.37 ^b | 5.91 ± 0.47 ^a | 4.94 ± 0.30 ^y |
| - Milk 30-d | 6.12 ± 0.37 ^b | 7.41 ± 0.46 ^a | 6.76 ± 0.29 ^x |
| - Milk 60-d | 5.86 ± 0.38 ^b | 7.22 ± 0.46 ^a | 6.54 ± 0.30 ^x |
| - Milk 90-d | 6.01 ± 0.40 ^a | 6.88 ± 0.47 ^a | 6.45 ± 0.30 ^x |
| Fat content, % | | | |
| - Colostrum | 4.38 ± 0.46 ^a | 5.31 ± 0.56 ^a | 4.85 ± 0.36 ^x |
| - Milk 30-d | 4.73 ± 0.47 ^a | 4.85 ± 0.58 ^a | 4.79 ± 0.37 ^x |
| - Milk 60-d | 5.36 ± 0.48 ^a | 5.27 ± 0.58 ^a | 5.32 ± 0.38 ^x |
| - Milk 90-d | 4.81 ± 0.52 ^a | 5.79 ± 0.60 ^a | 5.30 ± 0.40 ^x |
| Daily fat yield, g | | | |
| - Colostrum | 16.18 ± 3.32 ^b | 30.51 ± 4.24 ^a | 23.67 ± 2.69 ^y |
| - Milk 30-d | 28.92 ± 3.32 ^a | 36.13 ± 4.11 ^a | 32.52 ± 2.64 ^x |
| - Milk 60-d | 30.96 ± 3.39 ^b | 39.11 ± 4.11 ^a | 35.04 ± 2.66 ^x |
| - Milk 90-d | 29.50 ± 3.70 ^a | 37.38 ± 4.24 ^a | 33.44 ± 2.81 ^x |

Milk 30-d, Milk 60-d, and Milk 90-d refer to milk collected on the 30th, 60th, and 90th day of lactation, respectively.

Least squares means with different superscript letters (a, b) within a row are significantly different between Murrah and “Murrah × Carabao” crosses ($P < 0.05$).

Least squares means with different superscript letters (x, y) within the “total” column are significantly different between milk types ($P < 0.05$).

$$h/H = (C18:1n - 9 + PUFA) / [C12:0 + C14:0 + C16:0] \quad (4)$$

Fat Content Analysis

The fat content of each colostrum and milk sample was analyzed using the MilkoScan Mars (FOSS Analytical A/S, Hillerod, Denmark) that is based on Fourier-transformed infrared spectroscopy technology.

FA Analysis

Fat was extracted from colostrum and milk samples, following the method presented by Folch *et al.* (1957). Ten (10) mL of chloroform/methanol (2:1, v/v) was added to 1 mL of homogenized colostrum or milk sample and mixed for 1 min. Two mL of 1% NaCl solution was then added to induce phase separation and was remixed for about 30 s. Centrifugation was done at 10,000 rpm for 10 min. The lower organic phase was carefully collected

using a pipettor that was inserted through the protein disk interphase. The organic phase was finally transferred in a screw-capped glass test tube (16.5 × 105 mm).

The fatty acid methyl esters (FAMES) were prepared, following the rapid methanolysis/ methylation procedure (Ichihara and Fukubayashi 2010), using concentrated HCl. The lipid extract (organic phase) was placed in a screw-capped glass test tube sealed with a Teflon-lined cap. It was added with 3 mL of 8% methanolic HCl solution. Methylation was done in a water bath at 100 °C for 1 h. After cooling to room temperature, 1 mL of n-hexane and 3 mL of distilled water were added and mixed by shaking for 30 s. The sample in the glass tube was centrifuged for 5 min at 8000 rpm. The aqueous layer was removed while the hexane layer was transferred in 2 mL amber gas chromatography (GC) vials and purged with ultra-pure nitrogen gas for 20 s. The samples were stored in the refrigerator (−20 °C) prior to FA analysis by GC.

The FAMES were then quantified using a Shimadzu GC 2010 Plus capillary GC system (Shimadzu Corporation, Kyoto, Japan) that is equipped with a flame ionization detector and AOC-20i autosampler. It used a FAMEWax (USP G16) capillary column (30 m, 0.32 mm ID, and 0.25 μm film thickness, Restek Corporation, U.S.). The chromatographic conditions were as follows: the split ratio was 50:1, and the injection volume is 1 μL . The oven temperature was set to 125 $^{\circ}\text{C}$ and then increased to 240 $^{\circ}\text{C}$ at a rate of 3 $^{\circ}\text{C}/\text{min}$, where it was held for 5 min. The injection port was set to 230 $^{\circ}\text{C}$ while the detector temperature was set to 250 $^{\circ}\text{C}$. Hydrogen was used as a carrier gas at a flow rate of 40 mL/min, and nitrogen was used as make-up gas at a flow rate of 30 mL/min.

Quantitation of individual FAMES was based on their correction factors against the external standard. The FAME standard mix for 19 long-chained FAs was purchased from Sigma Aldrich and consisted of the grain FAME mix (CRM47801), AA (A3611), DHA (D2534), trans-vaccenic acid (V1131), and CLA (I6413). The FAME mix was analyzed at the start of each weekly batch of samples prepared for GC analysis. Accuracy was monitored by comparing the measured concentration of this FAME mix against its true concentration. Data (in triplicates) were processed using LabSolutions software. The FAs were identified by comparing their retention times with known FAME standards.

Statistical Analysis

Differences in colostrum or test-day milk yield (kg), fat content (%), and daily fat yield (g/d, *i.e.* test-day yield multiplied by fat content) among the milk types and between breeds (within the type of milk) were determined using ANOVA.

Pearson product-moment correlation coefficients among the individual FAs and their relationships with age at calving, parity, colostrum and test-day milk yield, and fat percentage were determined separately for colostrum and milk samples using the CORR procedure (SAS Institute Inc., Cary, NC).

The general least squares procedures for unbalanced data were used to examine the principal sources of variation affecting each FA in colostrum and milk. Statistical significance was set at $P < 0.05$. Only those significant fixed effects and covariates were included in the final linear models. Hence, the non-significant fixed effects of breed (*i.e.* Murrah and “Murrah \times Carabao” crosses) and covariate effect of age at calving (yr) were removed in the final model. The final mathematical model for all FAs was as follows: $y_{ijklm} = \mu + M\text{Type}_i + \text{Parity}_j + \text{Yield}_k + \text{PFat}_l + e_{ijklm}$, where y_{ijklm} is the percent area or the proportion of FA (g/100 g of total identified FAs) in buffalo colostrum

or milk, μ is the overall mean, $M\text{Type}_i$ is the fixed effect of the i^{th} type of milk (*i.e.* colostrum and milk collected on the 30th, 60th, and 90th day of lactation), Parity_j is the j^{th} covariate effect of parity, Yield_k is the k^{th} covariate effect of test-day yield (kg), PFat_l is the l^{th} covariate effect of fat percentage, and e_{ijklm} is the error term assumed to be normally distributed with the variance of errors as constant across observations.

The least-square means for each FA were used to compare the FA-based nutritional indices/ratios between milk types. Comparison of FA-based nutritional indices/ratios between breeds was based on the least-square means from the same mathematical model that included the effect of breed (*i.e.* Murrah and “Murrah \times Carabao” crosses) nested within the type of milk.

RESULTS

Amount of Fat Produced in Colostrum and Milk

Colostrum yield (4.9 kg) was significantly lower ($P < 0.05$) than daily milk yield (*i.e.* 6.4–6.8 kg obtained on the 30th, 60th, and 90th day of lactation, respectively) (see Table 1). While the fat percentage was similar ($P > 0.05$) between colostrum and milk, the total amount of milk fat produced per day was significantly lower ($P < 0.05$) for buffalo colostrum (23.7 g) than for milk (32.5–35.0 g). In practice, however, the processing of buffalo colostrum into high-value products in commercial quantities will be constrained by the small amount of colostrum that may be produced per calving, even in a year-round calving system that ensures a consistent supply of colostrum.

“Murrah \times Carabao” crosses had higher colostrum yield (5.9 kg *vs.* 4.0 kg) and daily milk yield (6.9–7.4 kg *vs.* 5.9–6.1 kg) than Murrah. “Murrah \times Carabao” crosses also produced more colostrum fat (30.5 g *vs.* 16.2 g) and more milk fat per day (36.1–39.1 g *vs.* 28.9–31.0 g) than purebred Murrah. However, the advantage of producing more milk fat in “Murrah \times Carabao” crosses over purebred Murrah, perhaps due to heterosis, will be of little importance in many local dairy farms that commonly raise a mix of purebred and crossbred buffaloes.

Correlations among Selected FAs

Among the major colostrum FAs with highest proportions and importance, oleic acid – a MUFA – was negatively correlated with SFAs [C12:0 ($r = -0.66$), C14:0 ($r = -0.74$), and C16:0 ($r = -0.43$)] and positively correlated with PUFAs [LA ($r = 0.37$) and AA ($r = 0.66$)]. Among the PUFAs, LA was positively correlated with ALA ($r = 0.37$) (see Table 2). Except for C20:4 n-6, all major FAs were not correlated with colostrum fat percentage ($P > 0.05$).

For milk FAs, oleic acid – a MUFA – was negatively correlated with SFAs [C12:0 ($r = -0.47$) and C14:0 ($r = -0.62$)] but positively correlated with C16:0 ($r = 0.20$) and C18:0 ($r = 0.18$). Oleic acid was positively correlated with PUFAs – LA ($r = 0.18$), ALA ($r = 0.32$), and AA ($r = 0.21$) (see Table 2).

Correlations of Selected FAs with Age at Calving, Parity, Colostrum or Test-day Milk Yield, and Fat Percentage

For the major FAs in colostrum, oleic acid was negatively correlated with age at calving ($r = -0.52$) and parity ($r = -0.63$) (see Table 3). Both C14:0 and C16:0 were not correlated with age at calving, parity colostrum yield, and fat percentage. In the case of omega-3 and omega-6 FAs, ALA was negatively correlated with parity ($r = -0.30$), while AA was positively correlated with age at calving ($r = 0.33$). The CLA, LA, and DHA were not correlated with age at calving, parity, colostrum yield, and fat percentage.

For milk FAs, oleic acid was negatively correlated with age at calving ($r = -0.18$) (see Table 3). C14:0 was positively correlated with age at calving ($r = 0.20$), parity ($r = 0.18$), and test-day yield ($r = 0.29$). C16:0 was positively correlated with age at calving ($r = 0.19$) and parity ($r = 0.19$). C18:0 was negatively correlated with test-day yield ($r = -0.29$). C12:0 was not correlated with age at calving, parity, and test-day yield. All major FAs were not correlated with milk fat percentage ($P > 0.05$). All PUFAs (CLA, LA, ALA, AA, and DHA) were negatively correlated with test-day yield ($r = -0.21, -0.26, -0.22, -0.18, \text{ and } -0.18$, respectively). ALA was also negatively correlated with age at calving ($r = -0.27$) and parity ($r = -0.27$).

Factors Affecting FA Composition

C12:0 was the most variable among the major SFAs with a coefficient variation (CV) of 27.50% – followed by C18:0 (CV = 19.47%), C14:0 (CV = 14.01%), and C16:0 (CV = 8.54%). By comparison, the CV for UFAs – oleic acid, LA, ALA, and CLA – were 23.19, 42.34, 20.07, and 70.06%, respectively (see Table 4).

All 19 FAs in the study were significantly affected ($P < 0.01$) by the type of milk. Higher parity was significantly associated ($P < 0.01$) with a decrease in the proportion of C14:0, C18:0, and ALA. The higher test-day yield was associated with an increase in C14:0 and a decrease in C18:0, LA, and CLA. A higher fat percentage was associated with an increase in C16:0.

Major FAs in Buffalo Colostrum

The three major FAs in buffalo colostrum – representing about 74% of total FAs – were C16:0 (32.3%), C18:1

n-9 (28.5%), and C14:0 (12.8%). These were followed by C18:0 and C12:0 with lower proportions of 5.4 and 4.4%, respectively (see Table 5). Colostrum contained smaller amounts of omega-6 FAs [*i.e.* LA (C18:2 n-6) and AA (C20:4 n-6) at 1.14% and 0.92%, respectively] and omega-3 FAs [*i.e.* ALA (C18:3 n-3) and DHA (C22:6 n-3 or DHA) at 0.36% and 0.26%, respectively]. Colostrum contained 0.64% CLA (C18:2 c9t11).

Major FAs in Milk Obtained on the 30th, 60th, and 90th Day of Lactation

The five major FAs in buffalo milk – representing about 82 to 84% of total FAs – were C16:0 (32.0–33.0%), C18:1 n-9 (18.0–18.8%), C14:0 (15.1–15.6%), C18:0 (8.7–9.7%), and C12:0 (7.8–8.2%) (see Table 5). Buffalo milk contained very small amounts of omega-6 FAs [*i.e.* LA and AA at 0.18–0.48% and 0.42–0.45%, respectively] and omega-3 FAs [*i.e.* ALA and DHA at 0.29–0.31% and 0.06–0.18%, respectively]. Milk contained 0.14–0.16% CLA. Except for myristoleic acid (C14:1 n-5) and palmitoleic acid (C16:1 n-7), all FAs in this study were not significantly different for milk obtained on the 30th, 60th, and 90th day of Lactation. Both C14:1 and C16:1 in milk collected on the 90th day of lactation (0.77 and 1.60%, respectively) was higher than on the 60th day (0.65 and 1.43%, respectively) and 30th day of lactation (0.64 and 1.41%, respectively).

DISCUSSION

Comparison of FAs between Milk Types

The SFAs were about 0.85 times lower in buffalo colostrum than in milk obtained on the 30th, 60th, and 90th day of lactation. On the other hand, the MUFAs were about 1.48–1.55 times higher in colostrum than in milk. The PUFAs were also 2.12–2.96 times higher in colostrum than in milk. Colostrum seems to provide a higher concentrated UFA diet for calves, although the metabolic needs in FAs of buffalo calves in the first days of life are unknown and a correlation with FA titers in colostrum is impossible (Coroian *et al.* 2013).

For the three major FAs with the highest proportions, C16:0 was similar in colostrum and milk. C14:0 was 0.82–0.85 times lower in colostrum than in milk. Oleic acid – the major MUFA, however – was 1.52–1.59 times higher in colostrum than in milk.

Omega-3 FAs (ALA and DHA) were 1.27–1.74 times higher in colostrum than in milk. Omega-6 FAs (LA and AA) were 2.22–3.38 times higher in colostrum than in milk.

Table 2. Pearson correlation coefficients among fatty acids in buffalo colostrum (upper right off-diagonals) and milk (lower left off-diagonals).

| FAs | SFA | | | | | | | | | | MUFA | | | | | | PUFA | | | | | |
|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|-------------|--------------|-------------|---------------|-------------|-------------|-------------|-------------|----|--|--|
| | 12:0 | 14:0 | 15:0 | 16:0 | 17:0 | 18:0 | 20:0 | 22:0 | 14:1 n-5 | 16:1 n-7 | 18:1 n-9 | 18:1 n-7 | 20:1 n-11 | 22:1 n-9 | 18:2 c9H11 | 18:2 n-6 | 18:3 n-3 | 20:4 n-6 | 22:6 n-3 | | | |
| SFA | | | | | | | | | | | | | | | | | | | | | | |
| C12:0 | - | 0.79 | ns | ns | -0.36 | ns | ns | ns | 0.33 | ns | -0.66 | ns | ns | ns | ns | -0.35 | ns | -0.39 | ns | ns | | |
| C14:0 | 0.77 | - | 0.43 | ns | -0.48 | -0.31 | 0.66 | ns | 0.66 | ns | -0.74 | ns | ns | ns | ns | ns | ns | -0.46 | ns | ns | | |
| C15:0 | ns | ns | - | 0.37 | ns | 0.36 | 0.31 | 0.60 | ns | 0.37 | -0.53 | 0.49 | 0.66 | 0.48 | ns | ns | 0.46 | -0.60 | ns | ns | | |
| C16:0 | -0.37 | ns | ns | - | -0.41 | ns | ns | ns | 0.42 | ns | -0.43 | ns | ns | ns | ns | ns | ns | ns | ns | ns | | |
| C17:0 | -0.60 | -0.71 | 0.26 | -0.19 | - | 0.49 | 0.50 | ns | -0.54 | ns | 0.36 | 0.38 | 0.41 | ns | ns | 0.40 | ns | ns | ns | ns | | |
| C18:0 | -0.34 | -0.42 | 0.20 | -0.38 | 0.56 | - | ns | 0.42 | -0.76 | -0.57 | ns | 0.41 | 0.60 | ns | ns | ns | ns | ns | ns | ns | | |
| C20:0 | -0.50 | -0.62 | ns | ns | 0.68 | 0.35 | - | 0.55 | ns | ns | ns | 0.43 | 0.36 | 0.34 | 0.50 | 0.44 | ns | ns | ns | ns | | |
| C22:0 | -0.23 | -0.24 | 0.40 | ns | 0.33 | 0.31 | 0.28 | - | ns | ns | ns | 0.49 | 0.79 | ns | ns | 0.45 | ns | ns | ns | ns | | |
| MUFA | | | | | | | | | | | | | | | | | | | | | | |
| C14:1 n-5 | 0.44 | 0.45 | -0.30 | ns | -0.54 | -0.74 | -0.40 | -0.28 | - | 0.48 | -0.39 | ns | ns | ns | ns | ns | ns | ns | ns | ns | | |
| C16:1 n-7 | -0.18 | -0.20 | -0.32 | 0.34 | ns | -0.59 | ns | ns | 0.21 | - | ns | ns | ns | ns | ns | ns | ns | ns | ns | ns | | |
| C18:1 n-9 | -0.47 | -0.62 | ns | 0.20 | 0.43 | 0.18 | 0.51 | ns | -0.52 | ns | - | ns | ns | ns | ns | 0.37 | ns | 0.66 | ns | ns | | |
| C18:1 n-7 | ns | ns | ns | ns | ns | ns | 0.21 | 0.21 | ns | ns | -0.24 | - | 0.64 | ns | 0.59 | 0.63 | ns | ns | ns | ns | | |
| C20:1 n-11 | -0.34 | -0.42 | 0.45 | ns | 0.42 | 0.49 | 0.45 | 0.82 | -0.31 | -0.25 | ns | 0.20 | - | ns | 0.37 | 0.55 | ns | ns | ns | ns | | |
| C22:1 n-9 | ns | 0.27 | -0.31 | ns | 0.26 | ns | 0.26 | - | ns | ns | ns | ns | ns | ns | | |
| PUFA | | | | | | | | | | | | | | | | | | | | | | |
| C18:2 c9H11 | ns | ns | - | ns | ns | ns | ns | ns | | |
| C18:2 n-6 | -0.58 | -0.23 | ns | ns | ns | 0.23 | 0.25 | ns | -0.44 | ns | 0.18 | 0.18 | 0.23 | ns | 0.60 | - | 0.37 | ns | ns | ns | | |
| C18:3 n-3 | -0.29 | -0.43 | 0.20 | ns | 0.31 | ns | 0.42 | 0.36 | -0.32 | ns | 0.35 | ns | 0.42 | 0.39 | 0.59 | 0.32 | - | ns | ns | ns | | |
| C20:4 n-6 | ns | -0.19 | ns | -0.30 | ns | ns | ns | -0.28 | -0.21 | ns | 0.53 | ns | ns | ns | 0.23 | 0.36 | ns | - | ns | ns | | |
| C22:6 n-3 | ns | ns | ns | ns | ns | ns | ns | - | | |

[ns] correlation coefficient is not significantly different from zero, $P > 0.05$

[Not in bold font] correlation coefficient is significantly different from zero, $P < 0.05$

[In bold font] correlation coefficient is highly significantly different from zero, $P < 0.01$

Table 3. Pearson correlation coefficients between individual FAs and age at calving, parity, colostrum/test-day milk yield, and fat percentage in buffalo colostrum and milk.

| FA | Colostrum | | | | Milk | | | |
|-------------|----------------|---------|-----------------|-------|----------------|---------|---------------------|-------|
| | Age at calving | Parity | Colostrum yield | % Fat | Age at calving | Parity | Test-day milk yield | % Fat |
| SFA | | | | | | | | |
| C12:0 | -0.31* | ns | ns | ns | ns | ns | ns | ns |
| C14:0 | ns | ns | ns | ns | 0.20* | 0.18* | 0.29** | ns |
| C15:0 | -0.51** | -0.49** | ns | ns | -0.25** | -0.27** | -0.24** | ns |
| C16:0 | ns | ns | ns | ns | 0.19* | 0.19* | ns | ns |
| C17:0 | ns | -0.31* | ns | ns | -0.29** | -0.31** | -0.18* | ns |
| C18:0 | -0.41** | -0.43** | ns | ns | ns | ns | -0.29** | ns |
| C20:0 | ns | ns | ns | ns | -0.19* | -0.19* | -0.19* | ns |
| C22:0 | -0.39** | -0.41** | -0.46** | ns | ns | ns | -0.24** | ns |
| MUFA | | | | | | | | |
| C14:1 n-5 | ns | ns | ns | ns | ns | ns | 0.23** | ns |
| C16:1 n-7 | 0.36* | 0.43** | ns | ns | ns | ns | ns | ns |
| C18:1 n-9 | -0.52** | -0.53** | ns | ns | -0.18* | ns | ns | ns |
| C18:1 n-7 | -0.40** | -0.44** | ns | 0.38* | ns | ns | ns | ns |
| C20:1 n-11 | ns | ns | ns | ns | -0.21* | -0.18* | -0.38** | ns |
| C22:1 n-9 | ns | ns | ns | ns | -0.28* | -0.30* | ns | ns |
| PUFA | | | | | | | | |
| C18:2 c9t11 | ns | ns | ns | ns | ns | ns | -0.21* | ns |
| C18:2 n-6 | ns | ns | ns | ns | ns | ns | -0.26** | ns |
| C18:3 n-3 | ns | -0.30** | ns | ns | -0.27** | -0.27** | -0.22* | ns |
| C20:4 n-6 | 0.33* | ns | ns | 0.35* | ns | ns | -0.18* | ns |
| C22:6 n-3 | ns | ns | ns | ns | ns | ns | -0.18* | ns |

[ns] correlation coefficient is not significantly different from zero, $P > 0.05$

[*] correlation coefficient is significantly different from zero, $P < 0.05$

[**] correlation coefficient is highly significantly different from zero, $P < 0.01$

Table 4. Mean square F tests for the effects of milk type and covariate effects of parity, test-day milk yield, and percent fat on the proportion of FAs (g/ 100 g of total identified FAs) in buffalo colostrum and milk.

| | Milk type | Parity | Test-day yield | Percent fat | Coefficient of variation (%) |
|-----------------------------|-----------|-------------------|-------------------|----------------|------------------------------|
| SFA | | | | | |
| C12:0, lauric acid | ** | ns | ns | ns | 27.50 |
| C14:0, myristic acid | ** | ns | *0.174 ± 0.082 | ns | 14.01 |
| C15:0, pentadecylic acid | ** | ** -0.020 ± 0.004 | ** -0.016 ± 0.005 | ns | 15.48 |
| C16:0, palmitic acid | ** | ns | ns | *0.188 ± 0.092 | 8.54 |
| C17:0, margaric acid | ** | ** -0.016 ± 0.004 | ns | ns | 23.07 |
| C18:0, stearic acid | ** | ** -0.147 ± 0.053 | ** -0.218 ± 0.064 | ns | 19.47 |
| C20:0, arachidic acid | ** | ns | ns | ns | 57.10 |
| C22:0, behenic acid | ** | * -0.002 ± 0.001 | ** -0.005 ± 0.001 | ns | 27.38 |
| MUFA | | | | | |
| C14:1 n-5, myristoleic acid | ** | ns | *0.016 ± 0.008 | ns | 26.42 |
| C16:1 n-7, palmitoleic acid | ** | *0.028 ± 0.011 | ns | ns | 20.78 |

| | Milk type | Parity | Test-day yield | Percent fat | Coefficient of variation (%) |
|--------------------------------|-----------|-----------------------|-----------------------|-------------|------------------------------|
| C18:1 n-9, oleic acid | ** | ns | ns | ns | 23.19 |
| C18:1 n-7, trans-vaccenic acid | ** | ns | ns | ns | 26.86 |
| C20:1 n-11, eicosenoic acid | ** | ** -0.004 ± 0.001 | ** -0.007 ± 0.001 | ns | 21.59 |
| C22:1 n-9, erucic acid | * | ns | ns | ns | > 100 |
| PUFA | | | | | |
| C18:2 c9 t11, CLA | ** | ns | * -0.019 ± 0.008 | ns | 70.06 |
| C18:2 n-6, LA | ** | ns | * -0.028 ± 0.010 | ns | 42.34 |
| C18:3 n-3, ALA | ** | ** -0.007 ± 0.002 | ns | ns | 20.07 |
| C20:4 n-6, AA | ** | ns | ns | ns | 33.81 |
| C22:6 n-3, DHA | ** | ns | ns | ns | > 100 |

[ns] no significant differences ($P > 0.05$); [*] significant differences ($P < 0.05$); [**] highly significant differences ($P < 0.01$)
Numbers in covariate columns are the regression coefficients and corresponding standard errors

Table 5. Least squares means of the proportion of FA (g/ 100 g of total identified FAs) and calculated FA groups plus FA-based nutritional indices/ratios for different milk types.

| | Colostrum | Milk 30-d | Milk 60-d | Milk 90-d |
|-------------------|----------------------------|---------------------------|----------------------------|---------------------------|
| C12:0 | 4.42 ± 0.32 ^b | 8.22 ± 0.30 ^a | 7.82 ± 0.31 ^a | 8.24 ± 0.33 ^a |
| C14:0 | 12.84 ± 0.34 ^b | 15.12 ± 0.32 ^a | 15.27 ± 0.32 ^a | 15.58 ± 0.34 ^a |
| C15:0 | 0.81 ± 0.02 ^b | 0.89 ± 0.02 ^a | 0.89 ± 0.02 ^a | 0.89 ± 0.02 ^a |
| C16:0 | 32.29 ± 0.45 ^{ab} | 32.03 ± 0.43 ^b | 32.78 ± 0.43 ^{ab} | 33.00 ± 0.46 ^a |
| C17:0 | 0.66 ± 0.02 ^a | 0.50 ± 0.02 ^b | 0.49 ± 0.02 ^b | 0.45 ± 0.02 ^b |
| C18:0 | 5.42 ± 0.26 ^c | 9.44 ± 0.25 ^a | 9.67 ± 0.25 ^a | 8.68 ± 0.27 ^b |
| C20:0 | 0.21 ± 0.01 ^a | 0.08 ± 0.01 ^b | 0.09 ± 0.01 ^b | 0.08 ± 0.01 ^b |
| C22:0 | 0.09 ± 0.01 ^b | 0.12 ± 0.00 ^a | 0.11 ± 0.00 ^a | 0.11 ± 0.01 ^a |
| C14:1 n-5 | 0.80 ± 0.03 ^a | 0.64 ± 0.03 ^b | 0.65 ± 0.03 ^b | 0.77 ± 0.03 ^{ab} |
| C16:1 n-7 | 1.88 ± 0.05 ^a | 1.41 ± 0.05 ^c | 1.43 ± 0.05 ^c | 1.60 ± 0.05 ^b |
| C18:1 n-9 | 28.49 ± 0.80 ^a | 17.96 ± 0.75 ^b | 18.80 ± 0.76 ^b | 18.37 ± 0.80 ^b |
| C18:1 n-7 | 0.11 ± 0.01 ^b | 0.15 ± 0.01 ^a | 0.14 ± 0.01 ^a | 0.16 ± 0.01 ^a |
| C20:1 n-11 | 0.13 ± 0.32 ^b | 0.19 ± 0.01 ^a | 0.18 ± 0.01 ^a | 0.17 ± 0.01 ^a |
| C22:1 n-9 | 0.06 ± 0.02 ^a | 0.02 ± 0.02 ^b | 0.02 ± 0.02 ^b | 0.02 ± 0.02 ^b |
| C18:2 c9 t11, CLA | 0.64 ± 0.03 ^a | 0.15 ± 0.03 ^b | 0.14 ± 0.03 ^b | 0.16 ± 0.03 ^b |
| C18:2 n-6, LA | 1.14 ± 0.04 ^a | 0.48 ± 0.04 ^b | 0.46 ± 0.04 ^b | 0.18 ± 0.04 ^c |
| C18:3 n-3, ALA | 0.36 ± 0.01 ^a | 0.31 ± 0.01 ^b | 0.31 ± 0.01 ^b | 0.29 ± 0.01 ^b |
| C20:4 n-6, AA | 0.92 ± 0.32 ^a | 0.45 ± 0.03 ^b | 0.42 ± 0.03 ^b | 0.43 ± 0.03 ^b |
| C22:6 n-3, DHA | 0.26 ± 0.05 ^a | 0.18 ± 0.05 ^{ab} | 0.07 ± 0.04 ^b | 0.06 ± 0.05 ^b |
| SFA | 56.74 | 66.39 | 67.11 | 67.03 |
| UFA | 34.78 | 21.93 | 22.60 | 22.20 |
| MUFA | 31.47 | 20.37 | 21.21 | 21.09 |
| PUFA | 3.31 | 1.56 | 1.39 | 1.12 |
| n-3 (ALA + DHA) | 0.61 | 0.48 | 0.38 | 0.35 |
| n-6 (LA + AA) | 2.06 | 0.93 | 0.88 | 0.61 |

| | Colostrum | Milk 30-d | Milk 60-d | Milk 90-d |
|----------------|-----------|-----------|-----------|-----------|
| PUFA/SFA ratio | 0.06 | 0.02 | 0.02 | 0.02 |
| LA/ALA ratio | 3.21 | 1.55 | 1.48 | 0.62 |
| n-6/n-3 ratio | 3.37 | 1.93 | 2.34 | 1.76 |
| IA | 2.53 | 4.59 | 4.50 | 4.66 |
| IT | 2.68 | 4.48 | 4.58 | 4.59 |
| HPI | 0.39 | 0.22 | 0.22 | 0.21 |
| h/H ratio | 0.64 | 0.35 | 0.36 | 0.34 |

Least squares means with different superscript letters within a row are significantly different ($P < 0.05$).

[Milk 30-d, Milk 60-d, and Milk 90-d] milk collected on the 30th, 60th and 90th day of lactation, respectively; [SFA] saturated fatty acids; [UFA] unsaturated fatty acids; [MUFA] monounsaturated fatty acids; [PUFA] polyunsaturated fatty acids; [LA] linoleic acid (C18:2 n-6); [ALA] α -linolenic acid (C18:3 n-3); [AA] arachidonic acid (C20:4 n-6); [DHA] docosahexaenoic acid (C22:6 n-3); [n-3] omega-3 fatty acids; [n-6] omega-6 fatty acids, [h/H ratio] hypocholesterolemic/ hypercholesterolemic ratio

The CLA was also 4.00–4.57 times higher in colostrum than in milk. The CLA is the best-known ruminant trans fats found in dairy fat and is widely recognized as an anticarcinogenic, antiatherogenic, anti-inflammatory, and weight-reducing substance (Rodriguez-Alcala *et al.* 2013; Agarwal and Gupta 2016; Whigham *et al.* 2007).

Comparison of Individual FAs with Other Breeds

FAs in buffalo colostrum. Except for AA, all FAs in this study were not significantly different between Murrah and “Murrah × Carabao” crosses ($P > 0.05$). AA in colostrum and milk fat was higher for Murrah (0.97 and 0.45–0.48%, respectively) than in “Murrah × Carabao” crosses (0.87 and 0.33–0.39%, respectively) (data not shown in Table 5). The AA is an essential omega-3 FA that helps keep cell membranes, especially those of nerve cells, healthy and permeable leading to stable nutrient supply in the brain and other parts of the body (Silva *et al.* 2019).

Colostrum from both Murrah and “Murrah × Carabao” crosses had lower SFAs, higher MUFAs, and higher PUFAs than colostrum from Romanian buffaloes reported by Coroian *et al.* (2013). In their study involving 10 SFAs and six UFAs, Coroian *et al.* (2013) also showed that some FAs in colostrum were affected by calving season, with higher concentrations of FAs possibly influenced by pasture nutrition during the summer season. They suggested that buffaloes can be milked starting with day four as the calves already benefit from a sufficient amount of colostrum and the surplus until Day 7 could easily be used for human nutrition. In terms of the major SFAs, colostrum from Murrah and “Murrah × Carabao” crosses contained lower C16:0 and C18:0, but higher C12:0 and C14:0 than those from Romanian buffaloes. Furthermore, Murrah and “Murrah × Carabao” crosses had higher oleic acid but lower LA than in the colostrum of Romanian buffaloes.

FAs in buffalo milk. The SFAs in milk from both Murrah and “Murrah × Carabao” crosses were similar to the SFAs reported for Kundi buffaloes in Pakistan (Talpur *et al.* 2007). The SFAs in milk in this study were however lower than those reported for Bulgarian Murrah (Ilieva *et al.* 2020), Nili-Ravi in Pakistan (Qureshi *et al.* 2015), Mediterranean in Italy (Pegolo *et al.* 2017), Anatolian water buffaloes in Turkey (Cinar *et al.* 2019), the Mediterranean in France (Menard *et al.* 2010), Bulgarian Murrah (Penchev *et al.* 2016), and Bhadawari in India (Kushwaha *et al.* 2018). The SFAs in milk in this study were higher than those reported for Murrah in Brazil (Fernandes *et al.* 2007) and the Mediterranean in Italy (Varricchio *et al.* 2007).

The MUFAs in milk from both Murrah and “Murrah × Carabao” crosses were similar to the MUFAs reported for Bhadawari in India (Kushwaha *et al.* 2018). The MUFAs in milk in this study were however lower than those reported for Nili-Ravi in Pakistan (Qureshi *et al.* 2015), Bulgarian Murrah (Penchev *et al.* 2016), Mediterranean in Italy (Pegolo *et al.* 2017), Mediterranean in Italy (Varricchio *et al.* (2007), the Mediterranean in France (Menard *et al.* 2010), Bulgarian Murrah (Ilieva *et al.* 2020), Kundi in Pakistan (Talpur *et al.* 2007), Anatolian water buffaloes in Turkey (Cinar *et al.* 2019), and Murrah in Brazil (Fernandes *et al.* 2007).

The PUFAs in milk from both Murrah and “Murrah × Carabao” crosses were similar to the PUFAs reported for Bhadawari in India (Kushwaha *et al.* 2018) and the PUFAs reported for the Mediterranean breed in France (Menard *et al.* 2010). The PUFAs in milk fat in this study were higher than the PUFAs reported for Anatolian water buffaloes in Turkey (Cinar *et al.* 2019). The PUFAs in milk in this study were however lower than those reported for Bulgarian Murrah (Penchev *et al.* 2016), Kundi in Pakistan (Talpur *et al.* 2007), Bulgarian Murrah (Ilieva *et al.* 2020), the Mediterranean in Italy (Pegolo *et al.* 2017),

Nili-Ravi in Pakistan (Qureshi *et al.* 2015), Murrah in Brazil (Fernandes *et al.* 2007), and the Mediterranean in Italy (Varricchio *et al.* 2007).

The oleic acid in milk from Murrah and “Murrah × Carabao” crosses was similar to the oleic acid reported for the Mediterranean in Italy (Pegolo *et al.* 2017). The oleic acid in milk in this study was higher than the oleic acid reported for Bhadawari in India in Turkey (Kushwaha *et al.* 2018). The oleic acid in milk fat in this study was, however, lower than those reported for the Mediterranean in France (Menard *et al.* 2010), Nili-Ravi in Pakistan (Qureshi *et al.* 2015), Bulgarian Murrah (Ilieva *et al.* 2020), Bulgarian Murrah (Penchev *et al.* 2016), Kundi in Pakistan (Talpur *et al.* 2007), Anatolian water buffaloes in Turkey (Cinar *et al.* 2019), and the Mediterranean in Italy (Varricchio *et al.* 2007).

The ALA in milk from Murrah and “Murrah × Carabao” crosses were similar to the ALA for the Anatolian water buffaloes in Turkey (Cinar *et al.* 2019) and the ALA reported for Bulgarian Murrah (Ilieva *et al.* 2020). The ALA in milk in this study were however slightly lower than those reported for Bhadawari in India (Kushwaha *et al.* 2018), Mediterranean in France (Menard *et al.* 2010), Kundi in Pakistan (Talpur *et al.* 2007), Nili-Ravi in Pakistan (Qureshi *et al.* 2015), Bulgarian Murrah (Penchev *et al.* 2016), Mediterranean in Italy (Pegolo *et al.* 2017), and the Mediterranean in Italy (Varricchio *et al.* 2007).

The LA in milk from Murrah and “Murrah × Carabao” crosses were similar to the LA reported for Bulgarian Murrah (Ilieva *et al.* 2020), Bhadawari in India (Kushwaha *et al.* 2018), and Nili-Ravi in Pakistan (Qureshi *et al.* 2015). The LA in milk in this study was, however, slightly lower than those reported for the Mediterranean in Italy (Pegolo *et al.* 2017), Bulgarian Murrah (Penchev *et al.* 2016), the Mediterranean in France (Menard *et al.* 2010), Kundi in Pakistan (Talpur *et al.* 2007), and the Mediterranean in Italy (Varricchio *et al.* 2007).

The CLA in milk from Murrah and “Murrah × Carabao” crosses were similar to those reported for Nili-Ravi in Pakistan (Qureshi *et al.* 2015) but slightly lower compared to the Mediterranean in Italy (Pegolo *et al.* 2017; Varricchio *et al.* 2007), Bulgarian Murrah (Ilieva *et al.* 2020), Kundi in Pakistan (Talpur *et al.* 2007), Mediterranean in France (Menard *et al.* 2010), Anatolian water buffaloes in Turkey (Cinar *et al.* 2019), and Murrah in Brazil (Fernandes *et al.* 2007).

FA-based Nutritional Indices/Ratios

The comparison of nutritional indices/ratios (related to cardiovascular health) among milk types and between breeds are shown in Tables 5 and 6, respectively.

PUFA/SFA ratio. The PUFA/SFA ratio is the commonly used index to assess the impact of diet on cardiovascular health (Chen and Liu 2020). The higher this ratio, the more positive the effect. This is because all PUFAs in the diet are known to depress low-density lipoprotein cholesterol

Table 6. Calculated values for FA groups and FA-based nutritional indices/ratios of colostrum and milk from Murrah and “Murrah × Carabao” (M×C) crosses.

| | Colostrum | | Milk 30-d | | Milk 60-d | | Milk 90-d | |
|-----------------|-----------|-------------|-----------|-------------|-----------|-------------|-----------|-------------|
| | Murrah | (M×C) cross |
| SFA | 56.51 | 56.87 | 65.83 | 67.34 | 66.66 | 67.84 | 67.13 | 66.96 |
| UFA | 34.94 | 34.89 | 22.81 | 20.45 | 23.25 | 21.55 | 22.35 | 21.84 |
| MUFA | 31.66 | 31.50 | 22.15 | 19.05 | 21.75 | 20.34 | 21.16 | 20.84 |
| PUFA | 3.28 | 3.39 | 1.66 | 1.40 | 1.50 | 1.22 | 1.19 | 1.00 |
| n-3 (ALA + DHA) | 0.58 | 0.66 | 0.53 | 0.41 | 0.38 | 0.36 | 0.35 | 0.35 |
| n-6 (LA + AA) | 2.09 | 2.06 | 0.98 | 0.85 | 0.97 | 0.73 | 0.67 | 0.52 |
| PUFA/SFA ratio | 0.06 | 0.06 | 0.03 | 0.02 | 0.02 | 0.02 | 0.02 | 0.01 |
| LA/ALA ratio | 3.14 | 3.28 | 1.57 | 1.52 | 1.54 | 1.38 | 0.52 | 0.47 |
| n-6/n-3 ratio | 3.61 | 3.12 | 1.83 | 2.06 | 2.52 | 2.02 | 1.93 | 1.48 |
| IA | 2.48 | 2.55 | 4.31 | 5.13 | 4.29 | 4.87 | 4.63 | 4.76 |
| IT | 2.69 | 2.63 | 4.27 | 4.87 | 4.47 | 4.76 | 4.60 | 4.60 |
| HPI | 0.40 | 0.39 | 0.23 | 0.19 | 0.23 | 0.21 | 0.22 | 0.21 |
| h/H ratio | 0.65 | 0.64 | 0.38 | 0.31 | 0.38 | 0.34 | 0.34 | 0.34 |

[Milk 30-d, Milk 60-d, and Milk 90-d] milk collected on the 30th, 60th and 90th day of lactation, respectively; [SFA] saturated fatty acids; [UFA] unsaturated fatty acids; [MUFA] monounsaturated fatty acids; [PUFA] polyunsaturated fatty acids; [LA] linoleic acid (C18:2 n-6); [ALA] α-linolenic acid (C18:3 n-3); [AA] arachidonic acid (C20:4 n-6); [DHA] docosahexaenoic acid (C22:6 n-3); [n-3] omega-3 fatty acids; [n-6] omega-6 fatty acids, [h/H ratio] hypocholesterolemic/ hypercholesterolemic ratio

and lower levels of serum cholesterol, whereas all SFAs contribute to high levels of serum cholesterol. However, not all PUFAs may positively affect the prevention of cardiovascular disease just as not all SFAs may contribute equally to serum cholesterol. For example, C20:4 n-6 (the major PUFA found in buffalo colostrum and milk) is known to increase high-density lipoprotein cholesterol level – which would neutralize some SFAs such as C12:0, C14:0, and C16:0 (Hanus *et al.* 2018). On the other hand, C14:0 and C16:0 – the major SFAs found in buffalo colostrum and milk – may increase total blood cholesterol levels (Zock *et al.* 1994) and, hence, widely thought to be one of the main causes of cardiovascular disease. In particular, the long-term consumption of SFAs in higher proportion to other healthy fats may increase the risk for cardiovascular disease (German and Dillard 2010). However, recent systematic reviews show a lack of association between saturated fat intake and cardiovascular disease (Siri-Tarino *et al.* 2010; de Souza *et al.* 2015). The SFAs, although not essential, also play numerous beneficial roles within the body. They are involved in lipogenesis, fat deposition, PUFAs bioavailability, and apoptosis (Legrand and Rioux 2010). Saturated fat also has protective effects against alcoholic liver disease (Kirpich *et al.* 2016).

In this study, the PUFA/SFA ratio was very low, *i.e.* 0.06: 1 for colostrum and 0.02: 1 for milk fat. The PUFA/SFA ratio in milk was similar for purebred Murrah (0.02–0.03: 1) and “Murrah × Carabao” crosses (0.01–0.02: 1). This suggests the very low amounts of PUFAs in relation to SFAs, and that healthier dairy products derived from either colostrum or milk regardless of the breed will need supplementation with PUFAs. For example, DHA (C22:6 n-3) may be added to help protect the cardiovascular system from the unhealthy effects of atherosclerotic lesions (Nacini *et al.* 2020).

LA/ALA ratio. LA and ALA are considered essential FAs because they cannot be synthesized by humans. Originally developed in guiding the manufacture of infant formula, the LA/ALA ratio describes the balance between LA (C18:2 n-6) and ALA (C18:3 n-3) as these PUFAs compete for the same desaturase and elongase enzymes, which they use to synthesize long-chain unsaturated FAs. The balance may be the most important factor, especially when long-chain PUFAs are not present in infant formulas (Chen and Liu 2020). When judging the nutritional value of baby food and infant formula, the LA/ALA ratio has a higher minimum reference value (within 5–15: 1). Tissues of adults have a lower rate of synthesis of n-3 long-chain PUFAs than those of infants, so the LA/ALA ratio in the diet does not have too much of an impact on adults.

In this study, LA/ALA ratio was low in colostrum (3.21: 1) and decreasingly lower in milk (0.62–1.55: 1) when

obtained from the 30th, 60th, until the 90th day of lactation. The LA/ALA ratio in milk was similar for purebred Murrah (0.52–1.57: 1) and “Murrah × Carabao” crosses (0.47–1.52: 1). Nonetheless, the low LA/ALA ratio implies the restricted use of buffalo colostrum or milk – regardless of breed – for infants.

n-6/n-3 ratio. The n-6/Omega-3 ratio is an important determinant of PUFAs and their coinciding effects on inflammatory diseases. The n-6 PUFAs (*i.e.* LA and AA) and n-3 PUFAs (*i.e.* ALA and DHA) are precursors to eicosanoids, which have important roles in the regulation of inflammation.

In general, eicosanoids derived from n-6 PUFA are pro-inflammatory, while eicosanoids derived from n-3 PUFA are anti-inflammatory. Increases in the n-6/n-3 ratio (as high as ~ 15:1 in the Western diet) are associated with greater metabolism of the n-6 PUFA compared with n-3 PUFA. This increase in the n-6/n-3 ratio coincides with increases in chronic inflammatory diseases such as nonalcoholic fatty liver disease, cardiovascular disease, obesity, inflammatory bowel disease, rheumatoid arthritis, and Alzheimer’s disease. Optimal dietary intakes of the n-6/n-3 ratio should be around 1–4: 1 (Patterson *et al.* 2012). A lower n-6/n-3 ratio (1–2: 1) may reduce the risk of many chronic diseases, although the optimal ratio may vary with the disease under consideration depending on the degree of severity of disease resulting from the genetic predisposition (Simopoulos 2002).

In this study, the n-6/n-3 ratio was lower (*i.e.* more balanced) for buffalo milk (1.8–2.3: 1) and highest (*i.e.* least balanced) for colostrum (3.4: 1). The n-6/n-3 ratio in milk obtained on the 30th, 60th, and 90th day of lactation was similar for Murrah (1.8–2.5: 1) and “Murrah × Carabao” crosses (1.5–2.1: 1).

IA. The IA originally proposed by Ulbricht and Southgate (1991), is a measure of the dietary contribution of some SFAs (C12:0, C14:0, and C16:0 except C18:0) that are pro-atherogenic (*i.e.* they favor the adhesion of lipids to cells of the circulatory and immunological systems). This is expressed in relation to all MUFAs and PUFAs that are anti-atherogenic (*i.e.* they inhibit the accumulation of fatty plaque and reduce the levels of phospholipids, cholesterol, and esterified FAs). Dietary fat with lower IA values, therefore, has greater health benefits (*i.e.* lower tendency to form fatty plaques in the arteries), as its consumption may reduce the risk of coronary heart disease (Chen and Liu 2020).

In this study, the IA was lower (*i.e.* greater health benefits) in colostrum (2.53) than in milk (4.50–4.66). The atherogenicity potential of milk from purebred Murrah (4.29–4.63) was slightly lower than in “Murrah × Carabao” crosses (4.76–5.13).

IT. The IT, developed by Ulbricht and Southgate (1991), is used to determine the dietary contribution of pro-thrombogenic SFAs (*i.e.* C12:0, C14:0, and C16:0) in relation to the anti-thrombogenic MUFAs and PUFAs. Consumption of foods or products with lower IT values (*i.e.* lower tendency to form clots in blood vessels) is beneficial for cardiovascular health (Chen and Liu 2020).

In this study, the IT was lower (*i.e.* more beneficial for cardiovascular health) in colostrum (2.68) than in milk (4.48–4.59). The differences in IT for milk of Murrah (4.27–4.60) and “Murrah × Carabao” crosses (4.60–4.87) were small.

HPI. The HPI as proposed by Chen *et al.* (2004) is the inverse of the IA. It is mainly used in research on dairy products such as milk and cheese with values commonly ranging from 0.16–0.68. Dairy products with a high HPI value are considered more beneficial to human health (Chen and Liu 2020).

In this study, the HPI was higher in buffalo colostrum (0.39) than in milk (0.21–0.22). The HPI values of milk from purebred Murrah (0.22–0.23) were slightly higher than in “Murrah × Carabao” crosses (0.19–0.21).

h/H ratio. The h/H ratio is a new index to assess the effect of dietary FA composition on cholesterol. First used by Mierlita (2018) in sheep milk, the h/H ratio characterizes the relationship between hypocholesterolemic FA (oleic acid and PUFA) and hypercholesterolemic FA (C12:0, C14:0, and C16:0). Compared with the PUFA/SFA ratio, the h/H ratio may more accurately reflect the effect of the FA composition on cardiovascular disease. Dietary fat with lower h/H values has greater health benefits. For dairy products, the range is 0.32–1.29: 1 (Chen and Liu 2020).

In this study, the h/H ratio was higher in buffalo colostrum (0.64: 1) than in milk (0.34–0.36: 1), suggesting that the use of colostrum may result in higher blood cholesterol levels. The h/H ratio milk from purebred Murrah (0.34–0.38: 1) was slightly higher than in “Murrah × Carabao” crosses (0.31–0.34: 1).

CONCLUSION

Buffalo colostrum seems to be more beneficial for cardiovascular health because of its lower IA and IT values and higher HPI than milk. However, raw milk had a more balanced n-6/omega-3 ratio and lower h/H ratio (*i.e.* related to lower blood cholesterol levels) than colostrum.

Milk from the Murrah breed may have slight advantages in terms of lower atherogenicity values and higher HPI values than milk from “Murrah × Carabao” crosses. On

the other hand, milk from “Murrah × Carabao” crosses had a slightly lower h/H ratio than milk from Murrah. The “Murrah × Carabao” crosses also produced more milk fat per day than Murrah.

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STATEMENT ON NO CONFLICT OF INTEREST

There is no conflict of interest between the authors and other people and organizations.

REFERENCES

- AGARWAL P, GUPTA R. 2016. A review on anticancer property of colostrum. *Res Rev – J Med Heal Sci* 5: 1–9.
- CHEN S, BOBE G, ZIMMERMAN S, HAMMOND EG, LUHMAN CM, BOYLSTON TD, FREEMAN AE, BEITZ DC. 2004. Physical and sensory properties of dairy products from cows with various milk fatty acid compositions. *J Agric Food Chem* 52: 3422–3428.
- CHEN J, LIU H. 2020. Nutritional indices for assessing fatty acids: a mini-review. *Int J Mol Sci* 21: 5695; doi:10.3390/ijms21165695
- CICHOSZ G, CZECHOT H, BIELECKA M. 2020. The anticarcinogenic potential of milk fat. *Ann Agric Environ Med* 27: 512–518.
- CINAR MU, OZSOY T, BEYZI SB, KALIBER M, KONCA Y. 2019. Milk and fatty acid composition of Anatolian water buffalo (*Bubalus bubalis*) from different provinces. *Buffalo Bull* 38: 107–118.
- COROIAN A, ERLER S, MATEA CT, MIREȘAN V, RADUCU C, BELE C, COROIAN CO. 2013. Seasonal changes of buffalo colostrum: physicochemical parameters, fatty acids and cholesterol variation. *Chem Cent J* 7: 40. doi: 10.1186/1752-153X-7-40

- DE SOUZA RJ, MENTE A, MAROLEANU A, COZMA AI, HA V, KISHIBE T, ULERYK E, BUDYLOWSKI P, SCHÜNEMANN H, BEYENE J, ANAND SS. 2015. Intake of saturated and trans unsaturated fatty acids and risk of all cause mortality, cardiovascular disease, and type 2 diabetes: systematic review and meta-analysis of observational studies. *BMJ* 351: h3978. doi: 10.1136/bmj.h3978.
- FERNANDES SAA, MATTOS W R S, MATARAZZO SV, TONHATI H, GAMA MAS, LANNA DPD. 2007. Total fatty acids in Murrah buffaloes milk on commercial farms in Brazil. *Ital J Anim Sci* 6: 1063–1066.
- FOLCH J, LEES M, SLOANE STANLEY GH. 1957. A simple method for the isolation and purification of total lipids from animal tissues. *J Biol Chem* 226: 497–509.
- GERMAN JB, DILLARD CJ. 2010. Saturated fats: a perspective from lactation and milk composition. *Lipids* 45: 915–923.
- HANUS O, SAMKOVÁ E, KRIZOVÁ L, HASONOVA L, KALA R. 2018. Role of fatty acids in milk fat and the influence of selected factors on their variability – a review. *Molecules* 23: 1636. doi: 10.3390/molecules23071636
- ICHIHARA K, FUKUBAYASHI Y. 2010. Preparation of fatty acid methyl esters for gas-liquid chromatography. *J Lipid Res* 51: 635–640.
- ILIEVA Y, IVANOVA S, PENCHEV P. 2020. Fatty-acid composition of buffalo milk under intensive and pasture farming. *J Cent Eur Agric* 21: 722–732.
- KIRPICH IA, MILLER ME, CAVE MC, JOSHI-BARVE S, MCCLAIN CJ. 2016. Alcoholic liver disease: update on the role of dietary fat. *Biomolecules* 6(1): 1. doi: 10.3390/biom6010001
- KUSHWAHA BP, SINGH S, MAITY SB, SINGH KK, MISRAAK, SINGH I. 2018. Milk fatty acid profile of Bhadawari buffaloes. *Indian J Anim Sci* 88: 868–870.
- LEGRAND P, RIOUX V. 2010. The complex and important cellular and metabolic functions of saturated fatty acids. *Lipids* 45: 941–946.
- MEHRA R, SINGH R, NAYAN V, BUTTAR HS, KUMAR N, KUMAR S, BHARDWAJA, KAUSHIK R, KUMAR H. 2021. Nutritional attributes of bovine colostrum components in human health and disease: a comprehensive review. *Food Biosci* 40. doi: 10.016/j.fbio.2021.100907
- MENARDO, AHMED S, ROUSSEAU F, BRIARD-BION V, GAUCHERON F, LOPEZ C. 2010. Buffalo *vs.* cow milk fat globule: size distribution, zeta-potential, compositions in total fatty acids and in polar lipids from the milk fat membrane. *Food Chem* 120: 544–551.
- MIERLITA D. 2018. Effects of diets containing hemp seeds or hemp cake on fatty acid composition and oxidative stability of sheep milk. *S Afr J Anim Sci* 48: 504–515.
- NAEINI Z, TOUPCHIAN O, VATANNEJAD A, SOTOUDEH G, TEIMOURI M, GHORBANI M, NASLI-ESFAHANI E, KOOHDANI F. 2020. Effects of DHA-enriched fish oil on gene expression levels of p53 and NF- κ B and PPAR- γ activity in PBMCs of patients with T2DM: a randomized, double-blind, clinical trial. *Nutr Metab Cardiovasc* 30: 441–447.
- O'CALLAGHAN TF, O'DONOVAN M, MURPHY JP, SUGRUE K, MANNION D, MCCARTHY WP, TIMLIN M, KILCAWLEY KN, HICKEY RM, TOBIN JT. 2020. Evolution of the bovine milk fatty acid profile – from colostrum to milk five days post parturition. *Int Dairy J* 104: 1–8.
- PATTERSON E, WALL R, FITZGERALD GF, ROSS RP, STANTON C. 2012. Health implications of high dietary omega-6 polyunsaturated fatty acids. *J Nutri Metab* 2: 539426. doi: 10.1155/2012/539426
- [PSA] Philippine Statistics Authority. 2020. Selected Statistics in Agriculture. Retrieved from https://psa.gov.ph/sites/default/files/2_SSA2020_final_signed.pdf
- PEGOLO S, STOCCO G, MELE M, SCHIAVON S, BITTANTE G, CECCHINATO A. 2017. Factors affecting variations in the detailed fatty acid profile of Mediterranean buffalo milk determined by 2-dimensional gas chromatography. *J Dairy Sci* 100: 2564–2576.
- PENCHEV P, ILIEVA Y, IVANOVA T, KALEV R. 2016. Fatty acid composition of buffalo and bovine milk as affected by roughage source – silage *versus* hay. *Emir J Food Agric* 28: 264–270.
- RODRIGUEZ-ALCALA LM, FONTECHA J, DE LA HOZ L, NUNES DA, SILVA VS, CARVALHO JE, BERTOLDO PACHECO MT. 2013. CLA-enriched milk powder reverses hypercholesterolemic risk factors in hamsters. *Food Res Int* 51: 244–249.
- QURESHI MS, MUSHTAQ A, JAN S, INYAT-UR-RAHMAN. 2015. Effect of age and lactation on milk fatty acid profile in dairy buffaloes. *Buffalo Bull* 34: 275–283.
- SAS INSTITUTE INC. 2009. SAS/STAT $\text{\textcircled{R}}$ 9.2 User's Guide, second edition.

- SILVA EGSO, RANGELAHN, MURMAML, BEZERRA MF, OLIVEIRA JPF. 2019. Bovine colostrum: benefits of its use in human food. *Food Sci Technol Campinas* 39(Suppl. 2): 355–362.
- SILVA GALDINO AB, RANGEL AHN, BUTTAR HS, NASCIMENTO MSL, GAVIOLI EC, OLIVEIRA RP, SALES DC, URBANO SA, ANAYA K. 2021. Bovine colostrum: benefits for the human respiratory system and potential contributions for clinical management of COVID-19. *Food Agric Immunol* 32: 143–162.
- SIMOPOULOS AP. 2002. The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomed Pharmacother* 56: 365–379.
- SIRI-TARINO PW, SUN Q, HU FB, KRAUSS RM. 2010. Meta-analysis of prospective cohort studies evaluating the association of saturated fat with cardiovascular disease. *Am J Clin Nutr* 91: 535–546.
- TALPUR FN, MEMON NN, BHANGER MI. 2007. Comparison of fatty acid and cholesterol content of Pakistani water buffalo breeds. *Pak J Anal Environ Chem* 8: 15–20.
- TIMLIN M, TOBIN JT, BRODKORB A, MURPHY EG, DILLON P, HENNESSY D, O'DONOVAN M, PIERCE KM, O'CALLAGHAN TF. 2021. The impact of seasonality in pasture-based production systems on milk composition and functionality. *Foods* 10: 607. doi: 10.3390/foods10030607
- ULBRICHT TLV, SOUTHGATE DAT. 1991. Coronary heart disease: seven dietary factors. *Lancet* 338: 985–992.
- VARRICCHIO M L, DI FRANZIA A, MASUCCI F, ROMANO R, PROTO V. 2007. Fatty acid composition of Mediterranean buffalo milk fat. *Italian J Anim Sci* 6: 509–511.
- WHIGHAM LD, WATRAS AC, SCHOELLER DA. 2007. Efficacy of conjugated linoleic acid for reducing fat mass: a meta-analysis in humans. *Am J Clin Nutr* 85: 1203–1211.
- ZOCK PL, DE VRIES JHM, KATAN MB. 1994. Impact of myristic acid versus palmitic acid on serum lipid and lipoprotein levels in healthy women and men. *Arterioscler Thromb* 4: 567–575.