Influence of Inoculation of Different Vesicular Arbuscular Mycorrhizal Fungi on Growth and Nutrient of Mungbean and Wheat

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An experiment was conducted under greenhouse conditions to evaluate the effects of different vescilarearbuseular mycorribal (VAM) fungl with and without Musscorie rock phosphate (MRP) in P-deficient natural non-disinfacted soil with mungbean (Vignar adiata L. Wilczek) and wheat (Tribum aesitivum L. mend Theil) as test crops. Root colonization by native VAM fungl was poor in uninoculated soils which was improved with inoculation. The MRP amendment stimulated root colonization. The plant blomass and nutrient uptake were greater in the presence of MRP. Highest plant blomass, N and Puptake was observed on numpdewn inoculated with Clamor Sackcularium in presence of MRP. In wheat clinicates, the Sack Clamor Sackcularium in presence of MRP. In wheat clinicates, the Sack Clamor Sackcularium in presence of MRP. In wheat clinicates, the Sack Clamor Sackcularium in presence of MRP. In wheat clinicates, the Sack Clamor Sackcularium in presence of MRP. In wheat clinicates, the Sack Clamor Sackcularium in presence of MRP. In wheat clinicates are the production, it and related typicate to the sack Clamor Sackcularium in presence of MRP. In wheat Clinicates are the production, and and Paculation and Clamor Sackcularium of the Clamor

Key words: Vesicular-arbuscular mycorrhizae; Mussoorie rock- phosphate; Vigna radiata, Triticum aestivum; Nutrient deficient soils

Vesicular-arbusoular mycorrhizal (VAM) fungi are known to play an important role in plant growth and nutrient uptake sepecially the supply of P (Kucey et al. 1998; Barea 1991; Solan 1991). The uptake of other nutrients such as zinc and copper are also reported to be increased by the inoculation with VAM fungi (Harley & Smith 1993; Fabre et al. 1990). The successful mycorrhizal association depends on the presence of host, fungus and appropriate proviousment.

VAM fungi have little specificity and any of them infect virtually any potential host plant (Smith & Gianinazzi-Pearson 1988; Gianinazzi 1991). However, VAM endophytes differ in their effectiveness which appears to be more dependent

on the specific host-endophyte combination Therefore, the selection of efficient VAM fungi is essential for inoculation of field crops. Most of the experiments regarding the selection of efficient VAM fungi in different crops have been conducted in sterile soils where competition from native VAM fungi is eliminated (Powell 1979; Weber et al. 1993). Limited information is available under unsterile natural conditions (Smith & Gianinazzi-Pearson 1988; Barea et al. 1988). The objectives of the present investigation were: (a) to find a suitable host-endophyte combination under natural nutrient limited soil conditions for mungbean and wheat and (b) to find out the effectiveness in promoting growth and N and P uptake by the different VAM endophytes in the presence or absence of Mussoorie rock phosphate (an insoluble P source).

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Materials and Methods

Experimental design

To study the effect of inocitation of different VAM ingl such as Glorine blocalistism. (That see ansu Gerd) (Gerd and Trappe), Glorines processes (Nocl and Gerd) (Gerd anna and Trappe) Glorinus sp. 88. Glorinus sp. 88. Glorinus sp. 88. Glorinus sp. 88. Glorinus sp. 89. See and Glorinus sp. 89. Glorinus sp. 89.

Soil

The soil used in the suby was taken from Haryana Agricultural University dyshorl dom. Hisara and it had the following characteristics city, 5.6%, sill 4.0%, specification of 9.4%, organic C, 0.10%, NahrQ-Q, overtactable, 9.25 ppm; total P, 0.001%; 4.9 c mole (p-) kg⁻¹ catione dramps expandly and pH 7.6. Four kgair dired scaled (2 mm) soil was placed in earther pots with a hole in the bottom to drawn out expanse.

Greenhouse conditions

Ten seeds were som at the depth of 2.5 cm and fair germination thronto 5 plants per 0.1 The pots being remination thronto 5 plants per 0.1 The pots were distributed at random in a greenhouse under antarial conditions. The mean ambient temperature through the control of the co

VAM inoculum and inoculation

VAM endophtes G ascibilatin (Thavater sensor agree) (and and Terap) and Glorum encosage (Nicol and Geori) (Gerdard mit page) and Glorum encosage (Nicol and Geori) (Gerdarman and Trappe), were obtained from D. J. Bagyars, Urriers lyd Agricultural Sciences, Bargaliere, India while Gornas ps. 85, Gloruse ps. 92 and Gloruses ps. 79 wes supplied by C. S. Dudeja, Deberton of Microsology, Harryana Agricultural Deberton of the Teraphology, Harryana Agricultural Deberton of the Sensor of Company of the Company of Company of the Company of Com

Rhizobium inoculation

Seeds of mungbean were inoculated with 72h old culture of Rhizobium sp. Vigna. S32 strain by putting one mt of culture (about 10° cells) over the each seed hole.

Rock phosphate

Mussonie rock phosphate (MRP) was obtained from M/S Pyrites, Phosphates and Chemisals Ltd., Noida, India as a 100-mesh size powder. It contained 8.1% total P. 0.056 mg ("water-soluble P. 0.18 mg/g on S M NaHCO, soluble P. 2.3 mg/g critic ael soluble P. and a pH (1.2 MRP-Water) 8.7. Phosphous in MRP is present as carbonate fluorapaths which consists of about 6.7% total rock phosphate and is of sedimentary origin.

Basal nutrients

MIPP was mixed well in each pot so as to suply do and 60 kg P.Q./ha in mungbean and wheat respectively. A basaid dose of nitrogen was applied to mungbean as ammonium suplines at 20 kg Nhai in case of wheat, N was added through urea to suply 120 kg Nhai in the upper 5 cm. The fertilizer doses were as per the recommendations based on earlier field experiments for each cross.

Plant sampling

Plant sampling for assessment of nodulation and intrigen fixation in unsuphase was desided by alter solving. Three pots was removed and analyzed for solving. Three pots was removed and analyzed for nodulation and introgenase activities (Indrag et al. 1968). The plants with other three pots for each retainments were harvested after 55 days to determine her not colonization, shoot and root biomass, and N and P uptake. Total N and P content of shoot and root were determined and uptake of N and P was calculated by "unsuphying with weight not can de aboot. Similarly," by "unsuphying with weight not can de aboot. Similarly," so the plant of the same frameter as for the municipean grow.

Parameters measured

At harvest, the dry matter yield was determined after drying the shoot and root to constant weight. The N and P content of root and shoot were determined.

Assessment of mycorrhizal infection

The root clearing and staining method of Phillips & Hayman (1970) was used for studying root colonization by VAM. After removal of shoot, the roots were thoroughly washed under tap water to remove the soil. Mycorrhizal colonization in cortical root tissue was estimated by examining microscopically stained root segments (1 cm long). Fity reot segments per pot were examined and the percentage of root segments colonized were scored.

Nutrient analysis

The microkjeldhahl method was used for N determination (Bremner 1980) and total P was determined by the method of John (1970).

Results and Discussion

VAM root colonization

The VAM root colonization ranged from 6.0 to 88.1% (table 1) and it was poer without VAM incoudation both in mungbean and inwheat. Incoudation with different VAM fungle of the significant increase in the degree of root colonization. Addition of MRP stimulated root colonization but we one-significant. Maximum root colonization [81.1%] was solvered with Maximum root colonization [81.1%] was solvered with Self-root allow in the significant increase of MRP. The interaction between rook phase levels and VAM incoulation was found to be enriporitional.

Dry matter yield

The results of inoculation with VAM fundi on dry matter production (shoot and root) by munqbean in absence and presence of MRP are shown in table 2. Both shoot and root dry matter production were improved due to inoculation with VAM fungi over inoculated control, the increase was significant even within treatments. The improvement in total plant biomass was more when MRP was added to the soil. In case of munabean the effectivity of Glomus fasciculatum in dry matter production was higher in presence of MRP (151%) as compared with that of without MRP (125%) over the inoculated control. The effect of two factor interaction of rock-phosphate with VAM fungi on total dry matter was nonsignificant. Similarly, in the case of wheat crop (table 3) the effectivity of Glomus sp. 88 in dry matter production was highest in the presence of MRP (153%) than in without MRP (116%) over the uninoculated control. Increased dry matter yield with different VAM inoculants over uninoculated control was significant.

Nutrient uptake

The results of the inoculation oldiflerent VAM fungi on P uptake of mungbean and wheat are depicted in tables 4 and 5. There was more P uptake in shoot as compared to root in both crops. Inoculation significantly improved P uptake both in the presence or absence of MAP. However, the P uptake was greater when the soil was amended with MRP. Amongst the different VAM fungl the maximum P uptake was observed unplants incoultated with Glomus fasciculatum in mungbean and Glomus sp. 88 in wheat and this was statistically sionificant.

Tables 6 and 7 show the effects of inoculation of significantly Mingi on Nuplake. VAM inoculation also significantly improved N uptake and maximum N uptake both in the absence and presence of MRP was observed on inoculation with Glomus fasciculatum in munobean and with Glomus so. 88 in wheat.

Nodulation and nitrogenase activity

There was no significant effect of VAM inoculation on nodule number, nodule dry weight and nitrogenase activity in mungbean (data not shown). Application of MRP did not improve nodulation and nitrogenase activity.

The results of the incoulation of different WAM fungi however duration in their orac obtinization ability. The different tensils of incord colonization of a host by different anotherly time gave an inclusion of a certain degree of conductivity and another included of a certain degree of (Glaininazz 1991). The enhancement of impromissal orac obtaination of the colonization of the Properties of trigonal symbolium. Similar increases in root colonization by WAM fungi in presence of rook Signed 1983, Too et al. 1996, Similar Assource 1999.

Both shoot and root biomass increased significantly in presence of VAM lungl which may be attributed due to the increased nutrient supply resulting form root colonization by VAM endophyte. The increase in plant growth on VAM inoculation has been earlier reported in various field crops (Schubert & Hayman 1990; Pearson & Jakobsen 1993; Rathore & Singh 1995).

The significant increase in P uptake by root and shoot in presence of different VAM I rung may be attributed due to increased physical explosition of the attributed due to increased physical explosition of the attributed due to increased physical explosition of the provided program of the presence of NIFP and VAM fungin accompaned to without NIFP may be due to better utilization of P from native DP as well as from root, phosphate (Bollan 1991. Fathero & Singh 1995, Singh 6 Nagoor 1996). Utilization of DP as well as from root, phosphate by VAM funging interpret in 1996. Utilization to solubilize P from NIFP (Singh & Nagoor 1996), in deather to solubilize P from NIFP (Singh & Nagoor 1996), in discipline and didnich, the presence of VAM fungin in histosphere harbour and microbial population within thay influence that the provided of the present that the provided production of th

Table 1. Effect of different VAM fungi on percent root colorization of mungbean and wheat with and without Mussoorie rock phosphate.

Treatments	Root Colonization (%)									
		Mungbean		Wheat						
	R _b	B	Mean		R	Mean				
Control (uninoculated)	6.6	6,0	6.31	8.3	13,3	10.8*				
Glomus sp. 85	73.4	80.3	76.9*	60.0	65.6	62.84				
Glomus sp. 88	62.2	67.8	65.04	73.3	80.3	76.8°				
Glomus sp. 92	74.3	81.3	77.8*	64.0	70.0	67.044				
Glomus sp. 97	66.6	75.5	71.1*	68.6	71.2	68.94				
Glomus mossees	68.8	73.3	71.11	65.6	68.3	68.9*				
Glomus lasciculatum	80.2	88.1	84.2*	62.6	69.3	66.0*				
Mean	61.7*	64.9		57.2*	62.6*					
LSD (P=0.005)										
Treatments (T)	4.6			4.8						
Rock Phosphate (R)	6.7			7.9						
TxR	NS			MS						

Treatment means followed by different letters are significantly different at 5% level of probability.

Table 2. Effect of different VAM fungi on dry matter yield of mungbean with and without Mussoprie rock phosphate

Trestments	Plot Colonization (%)										
	Mungbean			Wheat			Total				
	R,	R	Mean	P.	8	Mean	R.	R	Mean		
Uninoculated	6.6	6.0	6.31	8.3	13.3	10.8*	1.042	1.074	1.058*		
Glomus sp. 85	73.4	80.3	76.9°	609	65.6	62.8°	1,206	1.302	1.2534		
Glomus sp. 88	62.2	67.8	65.0°	731	80.3	76.89	1,190	1.318	1,254		
Glomus sp. 92	74.3	81.3	77.8°	649	70.0	67.0°°	1.157	1.321	1.239		
Glomus sp. 97	66.6	75.5	71.15	666	71.2	68.91	1.134	1,179	1.1579		
Glomus mosseae	68.8	73.3	71.1*	655	68.3	66.94	1,215	1.146	1.1819		
Glomus fascicutatum	80.2	88.1	84.2*	624	69.3	66.04	1,308	1.623	1,466		
Mean	61.74	64.5*		572	62.6°		1.179*	1.280*			
LSD (Pr0.05)							11110	11000			
Treatments (T)	0.039			O.037			0.070				
Rock Prosphate (R)	0.042			0.00			0.096				
TxR	0.051			NS			NS				
Treatment means folio	wed by d	ifferent le	itters are si	nolica náv di	Second at	S% found of	neobabilitu				

ignifica.mly different at 5% level of probability.

Table 3, Effect of different VAM fungi on dry matter yield of wheat with and without Musscorie rock phosphate.

Treatments	Dry matter yield (q pot ⁻¹)										
		Shoot		Shoot			Total				
	R _e	a	Mean	8	B	Mean	B.	B	Moan		
Uninoculated	0.627	0.637	0.632*	0.455	0.455	0.460*	1.082	1,102	1.092*		
Glamus sp. 85	0.695	0.758	0.727*	0.550	0.620	0.585	1.245	1.378	1.312		
Glomus sp. 88	0.708	0.928	0.8185	0.712	0.762	0.7374			1,555		
Glamus sp. 92	0.678	0.786	0.7320	0.578	0.604	0.591*	1.420	1.690			
Glomus sp. 97	0.685	0.717	0.702*				1.256	1.390	1.323*		
Glamus mossega	0.702	0.816		0.574	0.679	0.627*	1.260	1.395	1.328		
Glamus tasciculatum			0.759	0.578	0.624	0.6019	1.280	1.440	1.360 ^b		
Mean	0,688	0.724	0.696*	0.595	0.638	0.6179	1,264	1.362	1.313*		
LSD (P=0.005)	0.6814	0.7679		O.578*	0.627*		1.258*	1.394*			
Treatments (T)	0.031										
Rock Phosphate (R)				0.061			0.130				
Tx B	0.027			0.071			0.132				
	NS			0.092							
Treatment means folio	wed by d	Herent le	tters are sig	pilicenty di	Herent at	5% level of	prőbability.				

Table 4. Effect of different VAM fungi on P uptake by mungbean with and without Mussocrie rock phosphate.

Treatments	Puptake (µg por¹)										
	Shoot			Root			Total				
	R _c	R	Ikan	R,	R	Mean	R,	R	Mean		
Uninoculated	42	46	44*	28	32	304	70	78	744		
Glomus sp. 85	58	69	647	38	48	43°	96	117	1079		
Glomus sp. 88	52	61	570	37	58	48°	89	119	104 ^b		
Glomus sp. 92	56	81	69°	30	52	41 ^b	86	133	110°		
Glomus sp. 97	49	59	54*0	36	54	45°	85	103	949		
Glomus mosseae	52	78	85°	48	56	52 ⁶⁴	100	134	1179		
Glomus fasciculatum	73	88	81*	55	66	614	128	154	1416		
Mean	55*	69*		39*	52 ^b		93*	131 ^b			
LSD (P±0.005)											
Treatments (T)	16			8			30				
Rock Phosphate (R)	14			10			27				
T. D	A 10"			10			3.60				

Treatment means followed by different lettes a resignificantly different at 5% level of probability.

Table 5. Effect of different VAM fungi on P uplake by wheat with and without Mussoorie rock phosphate.

Treatments	Puptake (µg pot*)										
	Shoot			Root			Total				
	R,	В	llea.n	P _o	B	Mean	R,	R	Mean		
Uninoculated	32	37	35^	17	20	19*	49	57	53*		
Glomus sp. 85	69	89	790	49	67	58*	118	158	138*		
Glomus sp. 88	82	122	1029	66	89	78*	148	211	180*		
Glomus sp. 92	51	74	63**	40	52	469	91	126	1091		
Glomus sp. 97	46	72	59*	38	56	46*	82	138	1104		
Glomus mosseae	61	118	904	47	84	66**	108	202	1554		
Glomus fasciculatum	56	90	73°	36	78	579	92	168	1381		
Mean	57*	85*		42*	64 ^b		98*	1517			
LSD (P=0.05)											
Treatments (T)	18			14			25				
Rock Phosphate (R)	15			12			28				
T x R Freatment means folion	22			18			NS				

Table 6. Effect of different VAM fungion Nurplake by mungbean with and without Mussoorie rock phosphate.

Treatments				Nup	take (µg)	101°)			
	Shoot				Root			Total	
	R,	В	Meann	_R,	R	Mean	R ₂	R	Mean
Uninoculated	0.282	0.302	1292	0.185	0.192	0.189*	0.467	0.494	0.481*
Glomus sp. 85	0.435	0.662	1549	0.410	0.500	0.455°	0.845	1.162	1.0049
Glomus sp. 88	0.467	0.674	1571	0.431	0.512	0.4729	0.898	1.185	1.042*
Glomus sp. 92	0.442	0.728	1589	0.435	0.532	0.4849	0.878	1.260	1.069*
Glomus sp. 97	0.489	0.792	0.64 1**	0.463	0.564	0.5143	1.952	1,356	1.154*
Glomus massees	0.507	0.682	0.595*	0.495	0.625	0.560™	1.002	1,310	1,1564
Glomus fascicularum	0.585	0.868	0.727	0.53	0.684	0.607*	1.115*	1,552	1.334*
Mean	0.458*	0.673*		0.421*	0.516*		0.880*	1.189°	
LSD (Ps0.05)									
Treatments (T)	0.071			0.079			0.098		
Rock Phosphate (R)	0.074			0.052			0.075		
TxB	NS			NS			NS		

Treatment means followed by different letters are significantly different at 5% level of probability.

TxB

Table 7. Effect of different VAN furni on N uptake by wheat with and without Mussoorie rock phosphate.

Treatments				Nup	take (µ9 s	opt"			
	Short				Root		Total		
	R.	8	Mean	R,	R	Mean	R,	R	Mean
Uninoculated	0.232	0.25	0.245*	0.205	0.217	0.211*	0.437	0.474	0.456*
G/amys sp. 85	0.429	0.51	0.4717	0.382	0.416	0.399*	0.811	0.928	0.870
G/amus sp. 88	0.569	0.592	0.581°	0.534	0.563	0.549	1.103	1.555	1.129
Glomus Sp. 92	0.457	9.52	0.490°	0.391	0.484	0.4381	0.848	1.006	0.927*
Glomus sp. 97	0.428	0.48	0.458°	0.491	0.463	0.4329	0.829	0.952	0.891*
Glamus mossnar	0.506	0.56	0.53707	0.482	0.552	0.5174	0.988	1.120	1.0541
Glomus fasciculatum	0.463	0.53	0.500*	0.482	0.518	0.460*	0.865	1.054	0.9604
Mean	0.441*	0.497	-	0.400*	0.459°		0.840*	0.956*	
LSD (P=0.05)									
Treatments (T)	0.038			0.050			0.104		
Dark Phosphata (R)	0.021			0.039			0.981		

Treatment means lottowed by difer en letters are significantly different at 5% level of probability.

Our results did not show significant effect of incouration with himbiobium, and different VAM fungion nodule number, their dry weight and mitogenase activity of murphes. This is in contrasts other studies where VAM incoulation has been shown to enhance modulation and intropen fixation by rhanble (Barea et al. 1986, Fitter & Garbaye 1990). The significant al. 1996, Fitter & Garbaye 1990, The significant with the properties of the significant of the significant due to greater translocation of six N by VAM hyphae (Johnssen et al. 1991).

The results of the present sudy show that plants to bonass and nutrient uptake of numpleen and when crops improved by inoculation with YAM furnig and amendment of rock phosphate However, different YAM fungi performed differentially with host plant. YAM fungi performed differentially with host plant. Glomus fascioutatium and Glomus significant man of Glomus significant man of Glomus significant part of the presence of MIPD amendment.

References

- Barea JM. 1991. Vesicular-arbuscular mycorrhizae as modifiers of soil fertility. Adv. Soil. Sci. 15: 1-40.
- Barea JM, Azcon-Aguilar C & Azcon R. 1988. Interaction between myconhizal fungi and soil micoorganisms. In : NitrogenFixation by Legumes in Mediterranean Agriculture (eds. DP Beck & LA Masterson) pp. 153-162. Nijhtf., Dordrecht.
- Bolan NS. 1991. A critical review on the role of mycorrhizal fungi in the uptake of phosphorus by plants. Plant and Soil. 134; 189–297.
- Bremner JM. 1960. Determination Of nitrogen in soil by the Kieldahl method. J. Agrit. S ci. 55:11-33.

- Faber BA, Zasoki RJ, Buran RS & Urin K. 1990. Zinc uptake by corn as affected by vesicular-arbuscular mycorrhizae. Plant and Soil. 129: 121-130.
- Fitter AH & Garbaye J. 1994. Interactions between mycorrhizal fungi and other soil organisms. Plant and Soil 159: 123-132.
- George E, Marschner H & Jakobson I. 1991. Role of arbuscular mycorrhizal fungi in uptake of phosphous and nitrogen from soil. Critical Rev. Biotech. 15: 257-270.
- Gianinazzi S. 1991. Vesicular-arbuscular (endo) mycorrhizas : cellular, blochemical and genetic respects. Agric. Ecosyst. Environ. 35: 105-119. Hardy RWF. Holstein RD. Jackson EK & Burrs RC.
 - 1968. The acetylene ethylene assay for nitroger fixation: Laboratory and field evaluation. Pl. Physiol 43: 1185-1207.
- Harley JL & Smith SE. 1983. Mycorrhizal symbiosis. Academic Press, London. pp. 483.
- John MK. 1970. Colorimetric determination of phosphorus in soil and plant materials with ascorbic acid. Soil Sci. 109; 214-220.
 Johansen A, Jakobsen I & Jenson ES. 1992. Hyphal
- transport of 15 N-labelled nitrogen by a vesiculararbuscular mycombizal fungus and its effect on deplation of inorganic soil N. New Phytol. 122: 281-288. Kucey RMN, Janzen HH & Legett ME, 1969. Microbially
- mediated increase in plant available phosphorus. Adv. Agron. 42: 199-221.
- Pearson JN & Jakobson I. 1993. Symbiotic exchange of carbon and phosphorus between cucumber and three arbuscular mycorrhizal fungi. New Phytol. 124: 481-488.

- Phillips JW & Hayman DS, 1970. Improved procedure for clearing roots and staining parasitic and VA mycorrhizal fungi for rapid assessment of infection. Trans Rt Mycol Soc 55, 158-161.
- Powell Ct., 1979. Inoculation of white clover and ryegrass seed with mycorrhizal fungi. New Phytol. 83: 81-85.
- Rathore VP & Singh HP. 1995. Influence of vesiculararbuscular mycorrhizal fungi and phosphate on maize. J. Indian. Soc. Soil. Sci. 43: 207-210.
 - Schubert A & Hayman DS. 1990, Plant growth response to vesicular arbuscular mycorrhiza. XI: Effectiveness of different endophytes at different levels of soil phosphate. New Phytol. 103: 79-90.
 - Singh HP & Singh TA, 1993. The interaction of rock phosphate, Bradythizobium, vesicular-arbusoular mycorrhizae and phosphate-solubilizing microbes on soybean grown in a sub-Himalayan Mollisol. Mycorrhiza, 4: 37-43.

- Singh S & Kapoor KK, 1998. Effects of inoculation of phosphate-solubilizing microoganisms and an arbuscular mycorrhizal fungus on mungbean grown under natural soil conditions. Mycorrhiza 7: 249-253.
- Smith SE & Gianinazzi-Pearson V. 1988. Physiological interactions between symbionts in vesiculararbuscular mycorrhizal plants. Ann. Rev. Pl. Physiol. Pl. Mol Biol. 39: 221-244
- Toro M, Azcon R & Herrera R. 1996. Effects on yield and nutrition of mycorrhizal and nodulated Pueraria phaseoloides exerted by P-soubilizing rhizobacteria. Biol. Fertil. Soils. 21: 323-29.
- Weber E, Saxena MC, George E & Marschner H. 1993. Effect of vesicular-arbuscular mycorrhiza on vegetative growth and harvest index of chickpea grown in northern Syria. Field Crop Res. 32: 115-128.